

## AccuBase™ ELISA Kit

### PRODUCT INFORMATION

Product Name	Catalog Number	Size
AccuBase™ ELISA Kit	ACB-EE00B	96 T

### PRODUCT SPECIFICATION

Component Number	Component Name	Size	Component Description	Storage
ACB-EE00B-1	Pre-coated Plate	96 well	96-well plate pre-coated with anti-AccuBase™ monoclonal antibody	2-8°C
ACB-EE00B-2	300×AccuBase™ Protein Standard	3 vials	AccuBase™ protein standard (lyophilized). Reconstitute to a concentration of 3.6 µg/ml with 500µl ddH <sub>2</sub> O before use (300X)	
ACB-EE00B-3	20× Detection Antibody	750 µl/vial	Binds to AccuBase™ enzyme in samples	
ACB-EE00B-4	HRP Conjugate	15 ml/bottle	Binds to the detection antibody and catalyzes the reaction for color detection.	
ACB-EE00B-5	10× Assay Buffer	10 ml/bottle	For the diluting protein standard, sample, and antibody detection, Dilute to 1X before use.	
ACB-EE00B-6	20× Wash Buffer	30 ml/bottle	20× wash buffer containing PBS and Tween-20. Dilute to 1X before use	
ACB-EE00B-7	TMB	15 ml/bottle	Chromogenic substrate for HRP	
ACB-EE00B-8	Stop Solution	10 ml/bottle	0.5 M H <sub>2</sub> SO <sub>4</sub>	

### OTHER EQUIPMENT REQUIRED:

Microplate reader (full wavelength or with 450 nm filter), plate washer, incubating microplate shaker, pipette and pipette tips

### ASSAY PERFORMANCE

- (1) Detection range: 0.1875ng/ml-12 ng/ml
- (2) Sensitivity: 0.1 ng/ml
- (3) Accuracy: CV<10%

### STORAGE CONDITION AND EXPIRATION DATE

Store the kit components at 2-8°C. The kit is valid for 12 months from the production date. Reconstituted AccuBase™ standard is recommended to be stored at -80±10°C and avoids repeated freeze-thaw cycles.

### DETECTION PRINCIPLE

This kit uses sandwich ELISA to determine the concentration of AccuBase™ in the test sample. The capture AccuBase™ monoclonal antibody is pre-coated on the 96-well plate. AccuBase™ standard or test sample is added to the pre-coated 96-well plate and will bind to the capture antibody. The biotinylated detection antibody is then added to bind the AccuBase™-capture antibody complex, followed by the addition of streptavidin HRP conjugate to form the capture antibody-antigen-detection antibody-HRP complex.

The extra detection antibody and the HRP conjugate need to be washed off. The addition of TMB results in color changes. The amplitude of the color change is proportional to the amount of AccuBase™ that binds to the plate. The reaction is stopped by adding the stop solution and the absorbance is measured at 450nm. The sample AccuBase™ concentration is calculated from the AccuBase™ standards titration curve.

### OPERATING PROCEDURES

Equilibrate the kit to room temperature before use.

---

**Reagent Preparation:**

1. Prepare 1X Wash Buffer (Dilute 20X Wash Buffer with distilled H<sub>2</sub>O)
2. Prepare 1X Assay Buffer (Dilute 10X Assay Buffer with distilled H<sub>2</sub>O)
3. Prepare 1X Detection Antibody (Dilute the detection antibody with 1X Assay Buffer)

**Sample and standard Preparation**

4. **Preparation of AccuBase™ standards.** Add 500µL of ddH<sub>2</sub>O to one vial of standard. Dissolve the standard at room temperature for 20 min, mix gently (DO NOT vortex). The final concentration of the AccuBase™ standard is 3.6 µg/mL. Make the serial dilution of the AccuBase™ standard following the table below. Two-fold serial dilution of the AccuBase™ standard with 1X Assay Buffer for the titration curve in duplicates is recommended.

Standard	Concentration (ng/mL)	Dilution
A	12	3µL Standard in 900µL 1X Assay Buffer
B	6	1:2 dilution from Standard A with 1X Assay buffer
C	3	1:2 dilution from Standard B with 1X Assay buffer
D	1.5	1:2 dilution from Standard C with 1X Assay buffer
E	0.75	1:2 dilution from Standard D with 1X Assay buffer
F	0.375	1:2 dilution from Standard E with 1X Assay buffer
G	0.1875	1:2 dilution from Standard F with 1X Assay buffer
H (1X Assay Buffer)	0	N/A

**ELISA:**

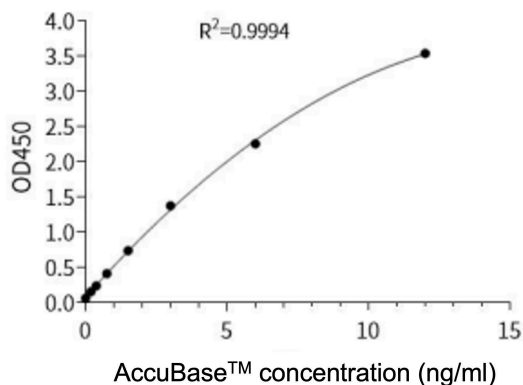
5. Equilibration: Take out the 96-well plate, seal unused strips, and immediately return to 4°C. Wash the plate with 300µL 1X wash buffer. Pat dry.
6. Incubation: Add the standards and samples to the 96-well plate (100µL per well). Incubate on a shaker (600 rpm) at 37°C for 1 hour.
7. Adding detection antibody: Wash the 96-well plate with 300µL 1X Wash Buffer 3 times. Pat dry and immediately add 1X detection antibody (100 µL per well). Incubate on the shaker (600 rpm) at 37°C for 1 hour.
8. Adding Streptavidin-HRP conjugate: Wash the 96-well plate with 300µL 1X Wash Buffer 3 times. Pat dry and add streptavidin-HRP conjugate (100 µL per well). Incubate on the shaker (600 rpm) at 37°C for 1 hour
9. Adding TMB Substrate: Wash the 96-well plate with 300µL 1X Wash Buffer 3 times. Pat dry and add 100 µL of TMB Substrate to each well. Incubate at 37°C for 10 minutes, protected from light.
10. Adding Stop Solution: Add 50µL of stop solution to each well and mix gently. Immediately read the OD value of each well at 450nm by a microplate reader. Reading the OD<sub>450</sub> values within 5 minutes after adding the stop solution is recommended.

**DATA ANALYSIS:**

1. Create a standard curve by plotting the standard concentrations on the x-axis against the OD<sub>450</sub> values on the y-axis of a scatterplot. If standards were run in duplicate or triplicate, use the average value. We recommended fitting the data with a 4-parameter logistic fit curve as the standard curve. Other methods such as linear and logarithmic methods may obtain better fitting results and may also be applicable, depending on the specific experimental needs.
2. Calculate the sample Accubase concentration by entering the sample OD<sub>450</sub> value into the equation of the standard curve. If you dilute the sample, multiply by the dilution factor. The lower limit of quantitation (LOQ) is 0.1875ng/mL. The sample should be further diluted and retested if the OD<sub>450</sub> of the sample falls above the valid linear range.

**EXAMPLE DATA:**

Standard (ng/mL)	OD450
12	3.540
6	2.260
3	1.380
1.5	0.742
0.75	0.415
0.375	0.243
0.1875	0.158
0	0.066



**NOTES:**

1. The microplate has detachable strips. Do not touch the bottom of the well while disassembling.
2. Do not leave the plate too long after each wash to avoid drying out.
3. 10X Assay Buffer and 20X Wash Buffer may precipitate at 4°C due to high salt concentration. The precipitant can be redissolved at room temperature.
4. Do not use this kit with components from other commercial kits, and do not mix components from different batches of kits. A standard curve must be prepared for each plate, and duplicates are recommended.
5. All reagents must be equilibrated to room temperature (18-25°C) before use. The TMB substrate should be warmed up to 37°C before use.
6. Make sure there is no liquid left in each well after each wash.
7. Using a plate washer can reduce the experimental error. For manual wash, it is recommended to soak the plate in 1X Wash Buffer for 1 minute after each addition.
8. The TMB substrate incubation needs to be protected from light and stopped within 10 minutes.
9. The Stop Solution contains sulfuric acid and may cause irritations on skin or eyes. Rinse immediately with plenty of water and seek medical assistance if necessary.